

Feature Article Commentary

Glial laminar cortical architecture matches metabolic demand

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The link between increased neural activity and cerebral blood flow is well established (Roy and Sherrington, 1890); yet, the mechanism by which blood flow is matched to neuronal activity both spatially and temporally is unclear. Based on their morphology, gray matter (protoplasmic) astrocytes are ideally situated to translate regional synaptic activity to hyperemia (increased flow) as they extend processes that envelop both synapses and vasculature (reviewed in Iadecola and Nedergaard, 2007). Indeed, evidence from *in vitro* and *in vivo* two-photon imaging has suggested that stimulus-induced elevations in astrocyte intracellular calcium are positioned to modulate vascular tone (Zonta *et al*, 2003; Takano *et al*, 2006; Mulligan and MacVicar, 2004; Winship *et al*, 2007). These local interactions are purported to occur at the astrocytic endfoot–blood vessel interface (reviewed in Iadecola and Nedergaard, 2007). Regionally, the spatial organization of astrocytes and vasculature should also have implications for matching blood flow to activity. While previous works have examined the interrelationships between astrocytes and microvasculature across cortical lamina, these studies have been conducted using histology and thus may not always faithfully represent the living tissue (White *et al*, 1981; Tsai *et al*, 2009). The present work of McCaslin *et al* (2011) has taken an important step to extend these findings in the live murine brain.

Using *in vivo* two-photon imaging, the authors found that the density of astrocytes showed distinct

peaks at 40 to 60 and 440 to 500 μm below the cortical surface, corroborating an earlier report (Tsai *et al*, 2009). Likewise, the density of capillaries mirrored that of astrocytes, reaching similar peaks at 30 to 40 μm and again at 500 μm ; moreover, the average distance between astrocytes and capillaries decreased with increasing cortical depth. This, together with evidence from Tsai *et al* (2009), who found that neuronal density peaks at 600 μm below the cortical surface, suggests that this columnar depth carries the highest metabolic load. Indeed, it has been recently reported that the fastest changes in dilation occur in diving arterioles and capillaries that are located at this cortical depth (Tian *et al*, 2010). Both experimental data (Tian *et al*, 2010) and models (Faraci and Heistad, 1990) have suggested that dilation of local arterioles is accompanied by temporally delayed dilation in upstream arteries. Gap junction-mediated communication between astrocytes has been implicated in the upward propagation of vasodilating signals from active neurons in the parenchyma to pial arteries (Xu *et al*, 2008). In support of a functional role for an astrocytic network in conducting hemodynamic signals, McCaslin *et al* (2011) confirm that a syncytium of astrocytes is in contact with all blood vessels below the glia limitans.

The laminar variations in astrocyte–vasculature interactions revealed by McCaslin *et al* (2011) may have important implications for the spatial and temporal matching of blood flow to activity within specific cortical layers. However, delineation of the relative contribution of distinct neuronal populations (reviewed in Cauli and Hamel, 2010) versus astrocytes to hemodynamic responses will require further exploration, perhaps with new optical tools to selectively regulate neuronal and astrocytic activity (Gradinaru *et al*, 2010).

Disclosure/conflict of interest

The author declares no conflict of interest.

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References

- Cauli B, Hamel E (2010) Revisiting the role of neurons in neurovascular coupling. *Front Neuroenergetics* 2:9
- Faraci FM, Heistad DD (1990) Regulation of large cerebral arteries and cerebral microvascular pressure. *Circ Res* 66:8–17
- Gradinaru V, Zhang F, Ramakrishnan C, Mattis J, Prakash R, Diester I, Goshen I, Thompson KR, Deisseroth K (2010) Molecular and cellular approaches for diversifying and extending optogenetics. *Cell* 141:154–65
- Iadecola C, Nedergaard M (2007) Glial regulation of the cerebral microvasculature. *Nat Neurosci* 10:1369–76
- McCaslin AFH, Chen BR, Radosevich AJ, Cauli B, Hillman EMC (2011) *In-vivo* 3D morphology of astrocyte-vasculature interactions in the somatosensory cortex: implications for neurovascular coupling. *J Cereb Blood Flow Metab* 31:795–806
- Mulligan SJ, MacVicar BA (2004) Calcium transients in astrocyte endfeet cause cerebrovascular constrictions. *Nature* 431:195–9
- Takano T, Tian GF, Peng W, Lou N, Libionka W, Han X, Nedergaard M (2006) Astrocyte-mediated control of cerebral blood flow. *Nat Neurosci* 9:260–7
- Tian P, Teng IC, May LD, Kurz R, Lu K, Scadeng M, Hillman EM, De Crespigny AJ, D'Arceuil HE, Mandeville JB, Marota JJ, Rosen BR, Liu TT, Boas DA, Buxton RB, Dale AM, Devor A (2010) Cortical depth-specific microvascular dilation underlies laminar differences in blood oxygenation level-dependent functional MRI signal. *Proc Natl Acad Sci USA* 107:15246–51
- Tsai PS, Kaufhold JP, Blinder P, Friedman B, Drew PJ, Karten HJ, Lyden PD, Kleinfeld D (2009) Correlations of neuronal and microvascular densities in murine cortex revealed by direct counting and colocalization of nuclei and vessels. *J Neurosci* 29:14553–70
- Roy CS, Sherrington CS (1890) On the regulation of the blood-supply of the brain. *J Physiol* 11:85–158.17
- White FP, Dutton GR, Norenberg MD (1981) Microvessels isolated from rat brain: localization of astrocyte processes by immunohistochemical techniques. *J Neurochem* 36:328–32
- Winship IR, Plaa N, Murphy TH (2007) Rapid astrocyte calcium signals correlate with neuronal activity and onset of the hemodynamic response *in vivo*. *J Neurosci* 27:6268–72
- Xu HL, Mao L, Ye S, Paisansathan C, Vetri F, Pelligrino DA (2008) Astrocytes are a key conduit for upstream signaling of vasodilation during cerebral cortical neuronal activation *in vivo*. *Am J Physiol Heart Circ Physiol* 294:H622–32
- Zonta M, Angulo MC, Gobbo S, Rosengarten B, Hossmann KA, Pozzan T, Carmignoto G (2003) Neuron-to-astrocyte signaling is central to the dynamic control of brain microcirculation. *Nat Neurosci* 6:43–50